



RESEARCH

Open Access

Accurate prediction of major histocompatibility complex class II epitopes by sparse representation via ℓ_1 -minimization

Clemente Aguilar-Bonavides^{1*†}, Reinaldo Sanchez-Arias^{2†} and Cristina Lanzas^{1,3}

*Correspondence:

clemen@nimbios.org

†Equal contributors

¹National Institute for Mathematical and Biological Synthesis, University of Tennessee, 37996-3410 Knoxville, TN, USA

Full list of author information is available at the end of the article

Abstract

Background: The major histocompatibility complex (MHC) is responsible for presenting antigens (epitopes) on the surface of antigen-presenting cells (APCs). When pathogen-derived epitopes are presented by MHC class II on an APC surface, T cells may be able to trigger a specific immune response. Prediction of MHC-II epitopes is particularly challenging because the open binding cleft of the MHC-II molecule allows epitopes to bind beyond the peptide binding groove; therefore, the molecule is capable of accommodating peptides of variable length. Among the methods proposed to predict MHC-II epitopes, artificial neural networks (ANNs) and support vector machines (SVMs) are the most effective methods. We propose a novel classification algorithm to predict MHC-II called sparse representation via ℓ_1 -minimization.

Results: We obtained a collection of experimentally confirmed MHC-II epitopes from the Immune Epitope Database and Analysis Resource (IEDB) and applied our ℓ_1 -minimization algorithm. To benchmark the performance of our proposed algorithm, we compared our predictions against a SVM classifier. We measured sensitivity, specificity and accuracy; then we used Receiver Operating Characteristic (ROC) analysis to evaluate the performance of our method. The prediction performance of MHC-II epitopes of the ℓ_1 -minimization algorithm was generally comparable and, in some cases, superior to the standard SVM classification method and overcame the lack of robustness of other methods with respect to outliers. While our method consistently favoured DPPS encoding with the alleles tested, SVM showed a slightly better accuracy when "11-factor" encoding was used.

Conclusions: ℓ_1 -minimization has similar accuracy than SVM, and has additional advantages, such as overcoming the lack of robustness with respect to outliers. With ℓ_1 -minimization no model selection dependency is involved.

Keywords: Peptide binding, Sparse representation, MHC class II, Epitope prediction, Immunoinformatics, Machine learning, Classification algorithms

Background

Pathogen peptide fragments are displayed on the surface of professional antigen presenting cells via the Major Histocompatibility Complex (MHC) class II. Such peptide fragments are known as epitopes. When helper T cells recognize epitopes bound to MHC-II, an adaptive immune response can be triggered for a specific pathogen. Computational prediction of MHC-II binding peptides can accelerate the development of vaccines and immunotherapies by identifying a narrow set of epitope candidates for further testing. Prediction of MHC-II epitopes is particularly challenging because the open binding cleft of the MHC-II molecule allows epitopes to bind beyond the peptide binding groove; therefore, the molecule is capable of accommodating peptides of variable length [1]. The binding core of the MHC-II is approximately nine amino acids long [2]; however, the complete epitope length can vary from 9 to 25 amino acids [3] and may even bind to whole proteins [4]. In addition, a successful computational prediction is based on sufficiently large set of high quality training data. Obtaining a large dataset for MHC-II epitope prediction can be difficult.

Machine-learning methods like artificial neural networks (ANNs) and support vector machines (SVMs) are classification techniques that have been successfully applied to predict MHC-II binding [5,6]. These methods, however, have some limitations. The biggest limitation of SVM lies in the choice of the kernel. The best choice of kernel for a given problem is still a research problem [7]. SVMs deliver a unique solution because the optimality problem is convex. On the other hand, ANNs have multiple solutions associated with local minima, which makes the method unrobust. The sparse representation (SR) approach proposed in this paper for peptide binding classification relies on the natural selective nature of the solution of an ℓ_1 -minimization problem [8]. This method overcomes the limitations of the machine-learning methods. Model selection is not necessary to differentiate between two classes; this contrasts with the need to test different kernel functions in the SVM approach when trying to find the best separating hyperplane with larger margin between classes. Furthermore, the use of the ℓ_1 norm promotes robustness in the method with respect to outliers in the data being used for classification [8], discarding bad training samples and allowing the handling of noisy data.

The ℓ_1 norm of a vector $x \in \mathbb{R}^n$ is defined as the sum of the absolute values of each of its components, i.e.,

$$\|x\|_1 = |x_1| + |x_2| + \dots + |x_n|. \quad (1)$$

Convex relaxation approaches based on the ℓ_1 norm have been proven to successfully promote sparse solutions (i.e. solutions with few nonzero elements) to linear system of equations with high probability. The work in the area of compressed sensing initiated in late 2004 by Emmanuel Candés, Justin Romberg and Terence Tao, and independently by David Donoho [8] encouraged the implementation of different fast solvers capable to find sparse solutions using the ℓ_1 norm as regularizer. Applications in science and technology have been successfully implemented with promising results in signal reconstruction, image processing, inverse problems, data analysis, among others. Finding sparse solutions has brought practical benefits such as the need of fewer antennas for remote sensing, fewer measurements needed in geophysical surveys, and more precise identification of genes.

recognition; therefore, this type of encoding is more informative. Secondly, property encoding mitigates the problem of flexible lengths. To test the reliability of property encoding, we used classical binary encoding and compared it against two property encoding methods, 11-factor encoding and divided physicochemical property scores (DPPS). The 11-factor encoding is calculated from physicochemical properties of amino acids as described by [12]. The properties were obtained from general physicochemical properties of amino acids and a number of properties identified in 3-D quantitative structure-activity relationship (QSAR) analysis [13]. The DPPS scheme was proposed by [14]. The DPPS descriptor was obtained by applying principal component analysis (PCA) to thousands of amino acid structural and property parameters. The resulting transformation yielded score vectors involving significant nonbinding properties of each of the 20 amino acids.

We represented every epitope of length n as a vector of 10 or 11 factors, corresponding to second and third encoding schemes, respectively, by adding to each position of the vector v_i the amino acid x_i 11-factor encoding or DPPS values in the following way:

$$v_i = \sum_{i=1}^n x_i, \tag{2}$$

Thus, every vector correlates directly with the physicochemical properties of amino acids, allowing the prediction of class II-peptide interaction. Additionally, we mitigated the problem of flexible lengths since every epitope is represented as a vector of size 10 or 11.

Classification via sparse representation

We applied the *selective nature* of sparse representation to perform classification. As presented in [8], ℓ_1 -minimization techniques provide a satisfactory method to solve sparse representation problems. We propose a classifier based on the solution of an ℓ_1 -minimization problem for classification. A supervised learning system performing classification is commonly called a *classifier*.

Formally, given an input dataset, $\mathbf{W} = \{\mathbf{w}_1, \dots, \mathbf{w}_n\}$, a set of labels/classes $\mathbf{T} = \{t_1, \dots, t_n\}$, and a training dataset $\mathbf{D} = \{(\mathbf{x}_i, t_i) : i = 1, \dots, n\}$, such that t_i is the label/class associated to the sample \mathbf{x}_i , a classifier is a mapping \mathcal{F} from \mathbf{W} to \mathbf{T} , assigning the correct label $t \in \mathbf{T}$ to a given input $\mathbf{w} \in \mathbf{W}$, that is, $\mathcal{F}(\mathbf{w}, \mathbf{D}) = t$.

Let us consider a *training data set* $\{(\mathbf{x}_i, t_i) : i = 1, \dots, n\}$, $\mathbf{x}_i \in \mathbb{R}^d$, $t_i \in \{1, 2, \dots, N\}$, where n is the number of samples and N the number of classes. The vector $\mathbf{x}_i \in \mathbb{R}^d$, represents the i th sample (for instance containing “gene expression” values, special features, etc), and t_i denotes its corresponding label (in our case, binding or non-binding). Assume that $d < n$, that is, the length of each sample is less than the number of elements in the training dataset.

The sparse representation problem is formulated as follows: For a testing sample $\mathbf{y} \in \mathbb{R}^d$, find the sparsest vector $\mathbf{c} = [c_1, c_2, \dots, c_n]^T$ such that

$$\mathbf{y} = c_1\mathbf{x}_1 + c_2\mathbf{x}_2 + \dots + c_n\mathbf{x}_n. \tag{3}$$

Equation (3) states that we express the vector \mathbf{y} as a linear combination of the collection $\{\mathbf{x}_1, \mathbf{x}_2, \dots, \mathbf{x}_n\}$. Using matrix algebra notation, equation (3) can be posed as the underdetermined linear system of equations

$$\mathbf{y} = \mathbf{A}\mathbf{c}, \tag{4}$$

where the matrix $\mathbf{A} \in \mathbb{R}^{n \times d}$ is constructed such that the j th column corresponds to sample \mathbf{x}_j , and the vector $\mathbf{c} = (c_1, \dots, c_n)^T$. Since we look for a sparse vector \mathbf{c} , equation (3) states that the test sample \mathbf{y} is a *linear combination of only a few training samples*. We are interested in the sparsest solution of the system of linear equations in (4). In order to find such a sparse solution, we solve the following ℓ_1 -optimization problem

$$\min_{\mathbf{c}, \mathbf{e}} \lambda \|\mathbf{c}\|_1 + \frac{1}{2} \mathbf{e}^T \mathbf{e} \tag{5}$$

subject to $\mathbf{A}\mathbf{c} - \mathbf{e} = \mathbf{y}$,

In [8], a novel optimization algorithm was proposed to solve problem (5) based on an iterative smooth convex relaxation methodology. One of the advantages of our formulation is that lack of robustness with respect to noise, missing data, and outliers can be overcome (a known property of the ℓ_1 norm is the regularization of an inverse problem). An additional advantage is that we do not need to care for model selection because the selective nature of the sparse representation captures the level of membership of a given input in one of the different classes. In the following section, we describe how to decide the class of a given input after obtaining its sparse representation. The approach consists of associating the nonzero components of \mathbf{c} with the columns of \mathbf{A} corresponding to those training samples that have the same class. First, let Ω_k denote the set of indices given by

$$\Omega_k = \{j : \text{training sample } \mathbf{x}_j \text{ has label } t_j = k\}. \tag{6}$$

Therefore,

$$\Omega_1 \cup \Omega_2 \cup \dots \cup \Omega_N = \{1, 2, \dots, n\}, \tag{7}$$

that is, the collection of sets of indices $\{\Omega_i\}_{i=1}^N$ forms a partition of the set $\{1, 2, \dots, n\}$, where n is the amount of samples available in the training dataset.

Then we define the *discriminant* functions by

$$g_k(\mathbf{y}) = \|\mathbf{y} - \mathbf{A}\mathbf{c}_k\|_2, \quad k = 1, \dots, N, \tag{8}$$

where $\mathbf{A}\mathbf{c}_k$ is defined by

$$\mathbf{A}\mathbf{c}_k = \sum_{j \in \Omega_k} c_j \mathbf{x}_j, \tag{9}$$

Notice that the function g_k in (8) measures the error obtained when the testing sample \mathbf{y} is represented with elements of the training set that have the same class k . Finally, we classify \mathbf{y} in the category with the smallest approximation error. That is, we compute

$$g_s(\mathbf{y}) = \min \{g_1(\mathbf{y}), g_2(\mathbf{y}), \dots, g_N(\mathbf{y})\}, \tag{10}$$

$\mathbf{t} = s,$

and conclude that the testing sample \mathbf{y} has label $\mathbf{t} = s$. In this manner, we identify the class of the test sample \mathbf{y} based on how effectively the coefficients associated with the training samples of each class recreate \mathbf{y} .

Algorithm 1 Classification Algorithm based on Sparse Representation (SR)

Goal: classify a given input \mathbf{y} in one of several categories $j = 1, \dots, N$.

Input: training matrix $A = [\mathbf{x}_1 | \mathbf{x}_2 | \dots | \mathbf{x}_n]$ and corresponding labels.

Output: label (class) $\hat{\mathbf{t}}$ of the given input sample \mathbf{y} .

Step 1: (optional) Normalize the columns of the matrix A to have unit norm.

Step 2: Solve the ℓ_1 minimization

$$\min_{\mathbf{c}, \mathbf{e}} \lambda \|\mathbf{c}\|_1 + \frac{1}{2} \mathbf{e}^T \mathbf{e} \quad \text{subject to } A\mathbf{c} - \mathbf{e} = \mathbf{y}.$$

Step 3: Calculate the N discriminant functions $g_k(\mathbf{y})$ as in (8)

Step 4: Find minimum discriminant value

$$g_s(\mathbf{y}) = \min \{g_1(\mathbf{y}), g_2(\mathbf{y}), \dots, g_N(\mathbf{y})\}.$$

Step 5: The input \mathbf{y} has label $\mathbf{t} = s$.

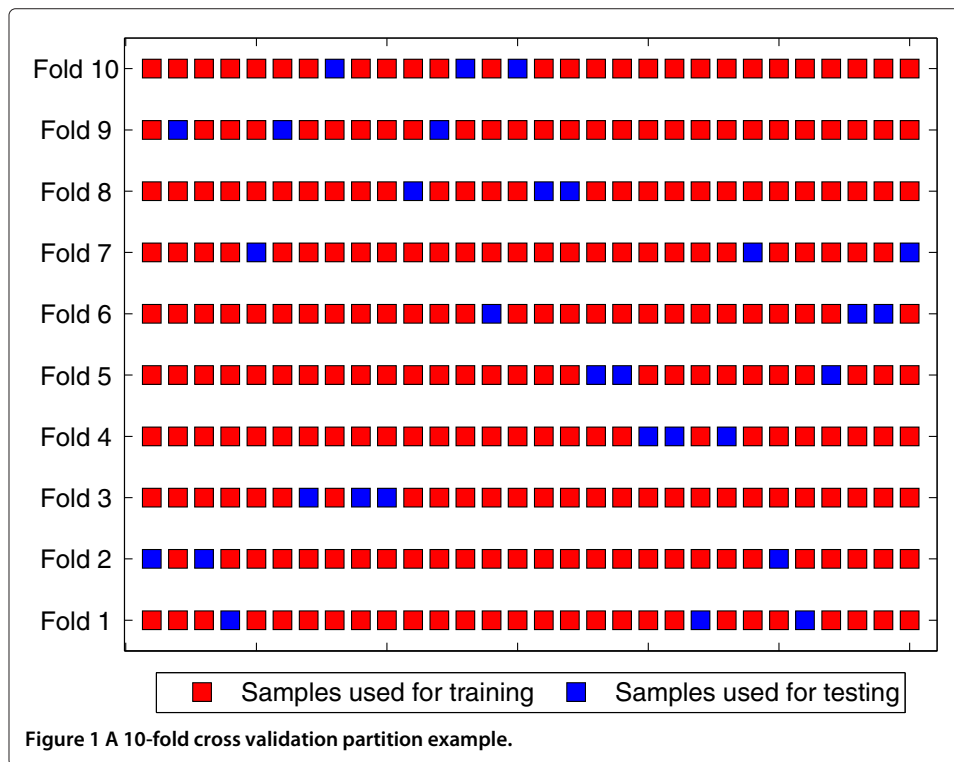
Support vector machines (SVM)

We compared the results of our proposed method for classification problems with the well known SVM strategy that has been commonly used in different pattern recognition and machine learning applications. SVMs are a set of related supervised learning methods that analyze data and recognize patterns commonly used for classification and regression analysis. The original SVM algorithm was proposed by Vladimir Vapnik and the current standard implementation was proposed by Corinna Cortes and Vladimir Vapnik, [15]. Standard SVM takes a set of input data and predicts for each given input which of two possible classes the input is a member of, which makes the SVM a non-probabilistic binary linear classifier. Intuitively, an SVM model is a representation of the samples as points in space, mapped so that the samples of the separate categories are divided by a clear gap that is as wide as possible.

Slow training is a possible drawback of SVM approaches because SVMs are trained by solving quadratic programming problems where the number of variables is equal to the number of samples in the training data set. When a large number of training data is available, the training process might turn slow. More information about the different strategies used in SVM for classification problems are described in [16]. Here we use the implementation of SVM available in MATLAB as part of the Statistics Toolbox, and report the results for the best possible setup (using radial basis functions) found after an appropriate parameter tuning stage (model selection).

Evaluation of method performance

To evaluate the prediction performance and robustness of our algorithm, we performed a 10-fold (n -fold) cross-validation. An illustration of the 10-fold cross validation partition process is shown in Figure 1. In the n -fold cross-validation, all the binding and non-binding epitopes were mixed and then divided equally into n parts, keeping the same distribution of binders and non-binders in each part. Then $n - 1$ parts were merged



into a training data set while the remnant was taken as a testing data set. This process was repeated 10 times and the average performance of n -fold cross-validation computed. We then measured sensitivity (Sn), specificity (Sp), accuracy (Acc) and Matthew's Correlation Coefficient (MCC) for every fold and then took the average (Avg) as shown in Table 2. In addition, we performed a ROC curves analysis using different IC_{50} thresholds.

Table 2 Average results for 10-fold cross-validation with ℓ_1 -minimization and SVM

	ℓ_1 -minimization					
	<i>H2-IA^d</i>			<i>HLA-DRB1*0101</i>		
	DPPS	11-factor	Binary	DPPS	11-factor	Binary
Avg. Sn (%)	89 (6)	86 (2)	81 (12)	97 (6)	97 (25)	97 (22)
Avg. Sp (%)	86 (10)	54 (9)	82 (10)	29 (10)	8 (22)	9 (21)
Avg. Acc. (%)	88 (4)	70 (7)	82 (8)	85 (8)	82 (12)	82 (15)
Avg. MCC	0.7558	0.4418	0.6441	0.3715	0.1	0
Time in secs	0.1650	0.1368	0.1368	1.5198	1.6831	1.9103
	SVM					
Avg. Sn (%)	78 (4)	81 (1)	78 (1)	83 (7)	97 (23)	100 (37)
Avg. Sp (%)	76 (10)	76 (2)	77 (6)	72 (12)	31 (20)	2 (37)
Avg. Acc. (%)	77 (3)	78 (6)	77 (8)	81 (7)	86 (10)	83 (19)
Avg. MCC	0.5559	0.5804	0.5694	0.4738	0.3951	0
Time in secs	0.1039	0.1268	0.1597	0.1770	0.2045	0.1928

Numbers in parenthesis indicate standard deviation.

We examined the association between cutoff value, encoding factor and method for every allele, after 10-fold cross-validation using logistic regression analysis. The results show that sensitivity and specificity are statistically associated with the three predictors in most cases, whereas no significant associations were seen in accuracy. Results shown in Table 3.

Results

Prediction accuracy

Techniques for predicting MHC binding include ANNs (NetMHCpan [6] and NN-Align [17]), position Specific Scoring Matrices (PSSMs) (RANKPEP [18]), and amino acid pairwise contact potentials as input vector for SVM (EpicCapo [10]). These methods have typical prediction accuracies of almost 70 – 90% [19]. Overall, our binding prediction accuracies are comparable to the reported 70 – 90% accuracies. Table 2 shows a comparison of the three encoding schemes with two alleles. While our method consistently favors DPPS encoding with the alleles tested, SVM shows a slightly better accuracy with 11-factor encoding. The experiments performed indicate that the physicochemical properties of amino acids are more informative in predicting MHC-II binding peptides. These results are also consistent with the MCC obtained for binary encoding, which yielded negative and zero scores on various occasions, implying that the predictions were not better than random predictions.

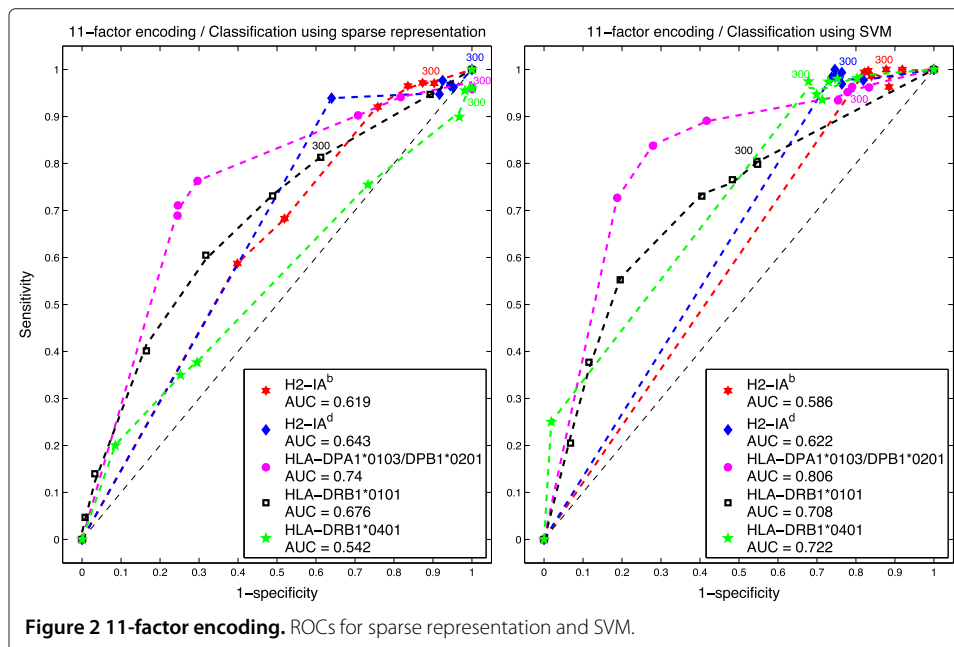
ROC curves analysis

We also applied ROC analysis to examine the performance of the ℓ_1 -minimization and SVM classifiers. An ROC graph is a plot with the false positive rate on the x axis (1 - specificity) and the true positive rate on the y axis (sensitivity). The point (0,1) is the perfect classifier: it classifies all positive cases and negative cases correctly. The point (0,0) represents a classifier that predicts all cases to be negative, while the point (1,0) is the

Table 3 Logistic regression p-values with cutoff value, encoding factor and predictive method as predictors

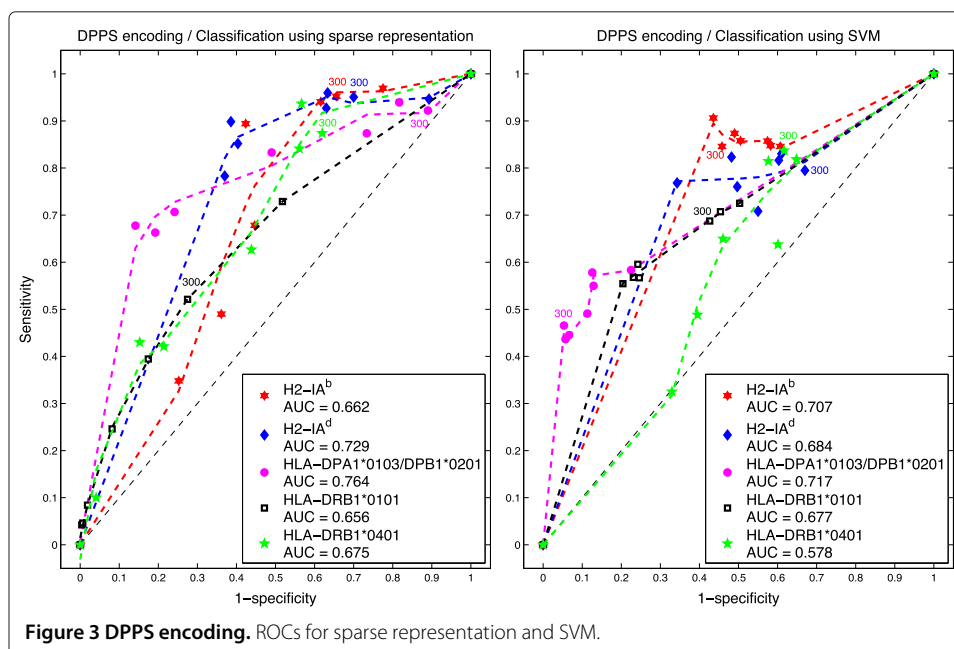
		Cutoff	Encoding	Method
H2-IA^b	Sn	***	***	*
	Sp	***	***	***
	Acc	***	NS	NS
H2-IA^d	Sn	NS	NS	***
	Sp	***	***	***
	Acc	NS	NS	NS
HLA-DPA1*0103/DPB1-0201	Sn	***	***	***
	Sp	***	***	***
	Acc	NS	NS	NS
HLA-DRB1*0101	Sn	***	***	**
	Sp	***	***	**
	Acc	***	NS	NS
HLA-DRB1*0401	Sn	***	***	**
	Sp	***	***	***
	Acc	NS	NS	NS

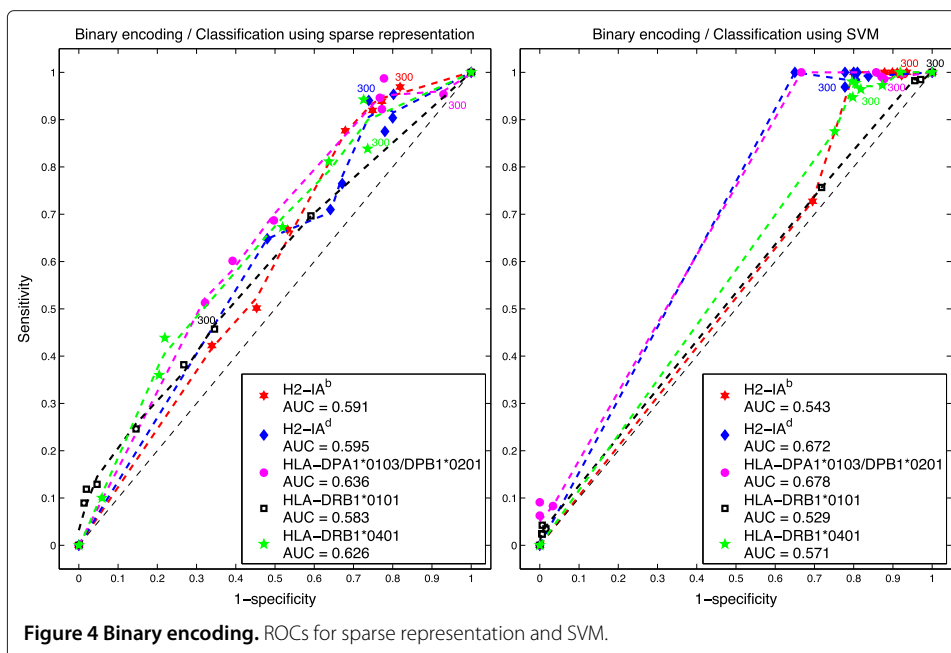
95% Confidence interval (CI) and P-values NS (No significant), *P < 0.05, **P < 0.01, ***P < .001.



classifier that is incorrect for all classifications. The ROC curves were calculated using the thresholds shown in Table 1 to distinguish binders from non-binders.

In Figures 2, 3 and 4 we present the corresponding ROCs for the sparse representation method and ROCs were calculated using different IC₅₀ cutoff values. The area under the ROC curve (AUC) provides a measure of overall prediction accuracy. An AUC value of 0.5 indicates random choice; while values close to 1 indicate excellent predictive capabilities of the method used. AUC values were computed using trapezoidal rule for numerical integration. With DPPS encoding, the ℓ_1 -minimization method for predicting





epitopes on H2-IA^d and HLA-DPA1*0103/DPAB1*0201 molecules rendered AUC values of 0.729 and 0.764, respectively, higher than any of the AUC of SVM for the same encoding scheme. However, with 11-factor encoding, the AUC obtained by SVM was 0.806 for molecule HLA-DPA1*0103/DPAB1*0201, a higher value than any AUC obtained by ℓ_1 -minimization. Tables 4, 5, 6, 7 and 8 show the values of sensitivity (Sn), specificity (Sp) and accuracy (Acc) for each of the IC₅₀ cutoff points, when using both the 11-factor and DPPS encoding schemes.

Discussion

Table 2 gives the results of our ℓ_1 -minimization algorithm and SVM predictions based on independent evaluation sets of three different epitope encoding methods. The

Table 4 11-factor and DPPS encoding for H2-IA^b

		11-factor encoding							
		IC ₅₀ cutoff	100	300	500	1000	5000	10000	15000
SR (ℓ_1)	Sn Avg (%)		100.00	97.05	97.15	96.51	92.00	68.28	58.72
	Sp Avg (%)		0.00	9.67	12.65	16.44	24.17	48.05	60.15
	Acc Avg (%)		90.00	81.82	79.41	73.58	62.38	57.49	59.60
SVM	Sn Avg (%)		99.40	98.58	99.74	100.00	100.00	99.56	96.28
	Sp Avg (%)		17.86	15.97	16.84	12.22	8.09	8.14	11.52
	Acc Avg (%)		91.94	84.11	82.32	74.84	59.80	50.82	45.31
		DPPS encoding							
SR (ℓ_1)	Sn Avg (%)		96.93	95.23	94.08	89.43	67.82	48.98	34.81
	Sp Avg (%)		22.50	34.44	38.41	57.65	55.34	63.87	74.77
	Acc Avg (%)		89.99	84.63	82.40	79.84	62.39	56.94	58.79
SVM	Sn Avg (%)		85.77	84.58	87.36	90.62	85.72	84.75	84.59
	Sp Avg (%)		49.50	54.22	51.01	56.44	42.57	41.77	39.28
	Acc Avg (%)		82.40	79.26	79.77	80.15	66.88	61.81	57.30

Table 5 11-factor and DPPS encoding for H2-IA^d

		11-factor encoding							
		<i>IC</i> ₅₀ cutoff	100	300	500	1000	5000	10000	15000
SR (ℓ_1)	Sn Avg (%)	100.00	100.00	94.74	96.58	93.90	96.19	97.65	
	Sp Avg (%)	0.00	0.00	8.33	5.56	36.00	4.76	7.50	
	Acc Avg (%)	90.00	80.00	75.80	77.00	70.48	70.22	68.80	
SVM	Sn Avg (%)	96.93	98.37	98.57	97.74	98.33	100.00	99.35	
	Sp Avg (%)	23.61	24.17	25.24	18.10	26.25	25.40	23.61	
	Acc Avg (%)	86.19	85.80	83.17	79.80	69.33	78.94	75.00	
		DPPS encoding							
SR (ℓ_1)	Sn Avg (%)	94.62	95.06	95.95	92.71	78.29	89.84	85.18	
	Sp Avg (%)	10.71	30.00	36.67	37.00	63.00	61.43	59.64	
	Acc Avg (%)	82.68	83.46	83.51	80.27	72.23	81.82	77.02	
SVM	Sn Avg (%)	70.82	79.50	81.63	76.03	83.05	76.83	82.32	
	Sp Avg (%)	45.00	33.00	39.67	50.33	39.00	65.71	51.79	
	Acc Avg (%)	67.33	71.39	72.98	70.08	65.30	73.73	72.63	

experiments performed with our method revealed binder average accuracies in the range of 70 – 88% for the alleles used, similar to those predictive accuracies reported elsewhere [19].

With DPPS encoding, the ℓ_1 -minimization method delivered higher AUC values of than any of the AUC of SVM for the same encoding scheme. However, with 11-factor encoding, the AUC obtained by SVM was higher than any AUC obtained by ℓ_1 -minimization. These results imply that different properties of amino acids are significant in the association process between the MHC-II molecule and the epitope, leading to higher performance in the prediction. This has a biological interpretation since nonbonding effects, such as electrostatic, van der Waals, hydrophobic interactions and hydrogen bond, play central roles in peptide-MHC interactions [14]. Hence, physico-chemical properties of amino acids should be considered when encoding epitopes for prediction. Since the ℓ_1 -minimization approach proposed here, where no model selection is involved, requires a more robust

Table 6 11-factor and DPPS encoding for HLA-DPA1*0103/DPB1*0201

		11-factor encoding							
		<i>IC</i> ₅₀ cutoff	100	300	500	1000	5000	10000	15000
SR (ℓ_1)	Sn Avg (%)	96.15	95.83	94.14	90.24	76.29	68.91	71.09	
	Sp Avg (%)	0.00	0.00	18.25	29.22	70.38	75.52	75.39	
	Acc Avg (%)	86.21	79.31	77.28	71.29	73.31	72.64	73.61	
SVM	Sn Avg (%)	96.19	96.32	95.20	93.44	89.10	83.78	72.69	
	Sp Avg (%)	16.67	21.00	22.14	24.57	58.29	72.09	81.24	
	Acc Avg (%)	88.02	82.83	78.72	72.21	73.59	77.06	77.69	
		DPPS encoding							
SR (ℓ_1)	Sn Avg (%)	93.93	92.22	87.37	83.29	70.67	66.28	67.76	
	Sp Avg (%)	18.33	11.00	26.67	51.00	75.86	80.78	85.85	
	Acc Avg (%)	85.82	77.69	73.64	73.30	73.31	74.67	78.44	
SVM	Sn Avg (%)	58.33	46.55	43.66	44.52	49.10	55.00	57.82	
	Sp Avg (%)	77.50	94.67	94.29	93.33	88.67	87.09	87.42	
	Acc Avg (%)	60.42	55.14	55.08	59.72	68.99	73.62	75.33	

Table 7 11-factor and DPPS encoding for HLA-DRB1*0101

		11-factor encoding							
		<i>IC</i> ₅₀ cutoff	100	300	500	1000	5000	10000	15000
SR (ℓ_1)	Sn Avg (%)	94.70	81.31	73.10	60.49	40.13	13.98	4.70	
	Sp Avg (%)	10.70	38.79	51.11	68.25	83.50	96.72	99.23	
	Acc Avg (%)	59.31	57.55	59.42	66.00	76.65	87.98	91.74	
SVM	Sn Avg (%)	80.46	79.84	76.55	73.05	55.24	37.70	20.51	
	Sp Avg (%)	45.39	45.22	51.70	59.53	80.45	88.48	93.20	
	Acc Avg (%)	65.67	65.24	62.66	64.64	76.46	83.11	87.43	
		DPPS encoding							
SR (ℓ_1)	Sn Avg (%)	72.89	52.12	39.46	24.60	8.41	4.61	4.24	
	Sp Avg (%)	48.24	72.52	82.56	91.84	98.13	99.33	99.59	
	Acc Avg (%)	62.51	63.52	66.27	72.36	83.96	89.32	92.03	
SVM	Sn Avg (%)	70.72	68.74	56.70	72.50	59.55	56.79	55.44	
	Sp Avg (%)	54.62	57.37	75.38	49.70	75.77	76.90	79.59	
	Acc Avg (%)	63.94	62.38	68.32	56.32	73.20	74.78	77.67	

way of presenting the information to the algorithm, in this case, we conclude that the DPPS encoding is more appropriate. This is because DPPS directly relates to peptide-MHC association. In Figure 5 we show the accuracy of the sparse representation and SVM methods when working with DPPS encoding for different *IC*₅₀ cutoff values. On the other hand, once the best choice of kernel has been obtained (model selection), SVM can handle less robust encoding schemes. We hypothesize that if more information is available in the encoding scheme of epitopes, our sparse representation algorithm could achieve higher performance.

Conclusions

The proposed ℓ_1 -minimization algorithm is able to produce accurate classification of MHC class II epitopes with sensitivity, specificity and accuracy to those from SVM approaches. We studied the algorithm performance for peptide binding classification and compared it with SVM for a collection of both human and mice alleles. Our methodology

Table 8 11-factor and DPPS encoding for HLA-DRB1*0401

		11-factor encoding							
		<i>IC</i> ₅₀ cutoff	100	300	500	1000	5000	10000	15000
SR (ℓ_1)	Sn Avg (%)	96.00	95.51	89.92	75.49	37.65	35.00	20.00	
	Sp Avg (%)	0.00	1.79	3.23	26.65	70.50	74.76	91.48	
	Acc Avg (%)	77.42	69.47	61.44	53.55	58.37	61.94	82.26	
SVM	Sn Avg (%)	97.40	97.46	98.10	93.56	94.70	98.00	25.00	
	Sp Avg (%)	27.04	32.14	19.73	28.63	30.08	24.76	98.15	
	Acc Avg (%)	84.28	78.95	72.30	64.45	53.86	48.39	86.45	
		DPPS encoding							
SR (ℓ_1)	Sn Avg (%)	93.65	87.37	84.10	62.68	42.12	43.00	10.00	
	Sp Avg (%)	43.33	38.06	43.91	56.15	78.58	84.76	95.93	
	Acc Avg (%)	84.23	73.53	70.94	59.70	65.18	71.29	84.84	
SVM	Sn Avg (%)	83.68	81.45	81.85	64.97	48.82	63.75	32.50	
	Sp Avg (%)	38.33	42.36	35.15	53.96	60.47	39.88	67.04	
	Acc Avg (%)	75.11	70.40	66.33	60.00	56.26	47.58	62.58	

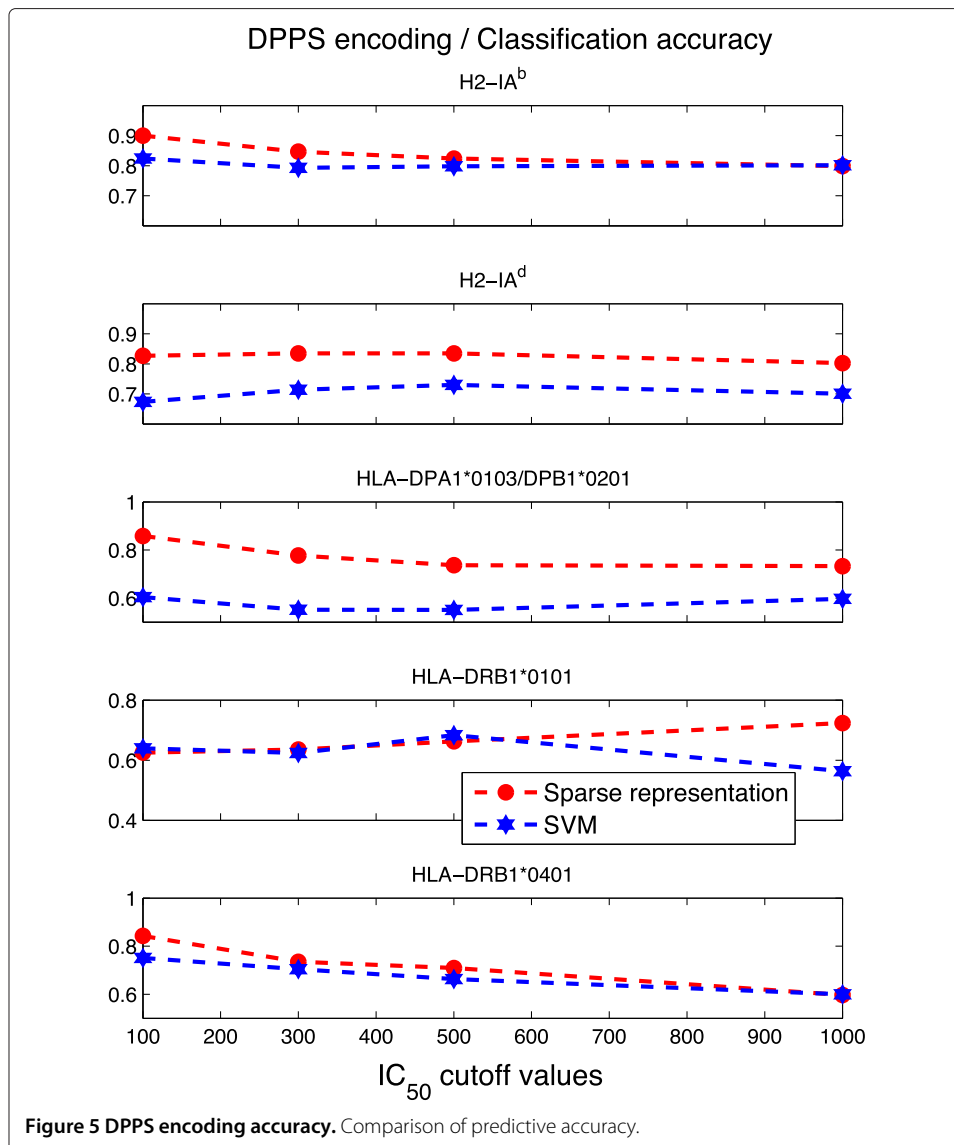


Figure 5 DPPS encoding accuracy. Comparison of predictive accuracy.

relies on the natural selective nature of sparse representation in order to perform classification wherein no model selection is involved; with regards to robustness to outliers, our classification enabled us to discard bad training samples and handle noisy data [8,20]. This contrasts with the need to test different kernel functions in the SVM approach when trying to find the best separating hyperplane with a larger margin between classes. Our methodology involves a very simple learning stage and the use of an ℓ_1 -minimization solver first proposed in [8]. For the set of alleles studied in this work, we found the DPPS encoding scheme to be efficient in conjunction with the proposed methodology for peptide binding classification.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

CAB conceived the idea of testing the algorithm for epitope prediction, gathered the data and prepared the datasets used during the research presented in the paper, and performed the experiments shown in the Results section. RSA

conceived the algorithm proposed in the manuscript, wrote the computer programs implementing the method, and designed the figures interpreting the results. CL provided ideas about how to test the predictive accuracy of the algorithm. CAB, RSA, and CL collaborated in the writing of the manuscript, and read and approved the final version.

Acknowledgements

This work was conducted while CAB was a postdoctoral fellow at the National Institute for Mathematical and Biological Synthesis, an Institute sponsored by the National Science Foundation, the U.S. Department of Homeland Security, and the U.S. Department of Agriculture through NSF Awards No. EF-0832858 and No. DBI-1300426, with additional support from The University of Tennessee, Knoxville. The authors thank Misty Bailey from the University of Tennessee for providing editorial comments.

Author details

¹National Institute for Mathematical and Biological Synthesis, University of Tennessee, 37996-3410 Knoxville, TN, USA.

²Department of Applied Mathematics, Wentworth Institute of Technology, 02115 Boston, MA, USA. ³Department of Biomedical and Diagnostic Sciences, University of Tennessee, 37996-3410, Knoxville, TN, USA.

Received: 3 March 2014 Accepted: 25 October 2014

Published: 4 November 2014

References

1. Wang P, Sidney J, Dow C, Mothé B, Sette A, Peters B: **A systematic assessment of MHC class II peptide binding predictions and evaluation of a consensus approach.** *PLoS Comput Biol* 2008, **4**(4):e1000048. doi:10.1371/journal.pcbi.1000048.
2. Lundegaard C, Lund O, Kesmir C, Brunak S, Nielsen M: **Modeling the adaptive immune system: predictions and simulations.** *Bioinformatics* 2007, **23**:3265–3275.
3. Patronov A, Dimitrov I, Flower D, Doytchinova I: **Peptide binding prediction for the human class II MHC Allele HLA-DP2: a molecular docking approach.** *BMC Struct Biol* 2011, **11**:32.
4. Nielsen M, Lundegaard C, Worning P, Hvid C, Lamberth K, Buus S, Brunak S, Lund O: **Improved prediction of MHC class I and class II Epitopes using a novel Gibbs sampling approach.** *Bioinformatics* 2004, **20**:1388–1397.
5. Bhasin M, Raghava G: **SVM based method for predicting HLA-DRB1*0401 binding peptides in an antigen sequence.** *Bioinformatics* 2004, **20**:421–423.
6. Nielsen M, Justesen S, Lund O, Lundegaard C, Buus S: **NetMHCIIpan-2.0 - Improved Pan-Specific HLA-DR predictions using a novel concurrent alignment and weight optimization training procedure.** *Immunome Res* 2010, **6**:9.
7. Wu KP, Wang SD: **Choosing the kernel parameters for support vector machines by the inter-cluster distance in the feature space.** *Pattern Recognit* 2009, **42**:710–717.
8. Sanchez-Arias R: **A convex optimization algorithm for sparse representation and applications in classification problems.** *PhD thesis.* The University of Texas at El Paso; 2013.
9. Nielsen M, Lundegaard C, Blicher T, Peters B, Sette A, Justesen S, Buus S, Lund O: **Quantitative predictions of peptide binding to any HLA-DR molecule of known sequence: NetMHCIIpan.** *PLoS Comput Biol* 2008, **4**:e1000107. doi:10.1371/journal.pcbi.1000107.
10. Saethang T, Hirose O, Kimkong I, Tran V, Dang X, Nguyen L, Le T, Kubo M, Yamada Y, Satou K: **EpicCapo: Epitope prediction using combined information of amino acid pairwise contact potentials and HLA-peptide contact site information.** *BMC Bioinformatics* 2012, **13**:313.
11. Kim Y, Ponomarenko J, Zhu Z, Tamang D, Wang P, Greenbaum J, Lundegaard C, Sette A, Lund O, Bourne P, Nielsen M, Peters B: **Immune epitope database analysis resource.** *Nucleic Acids Res* 2012, **40**:525–530.
12. Liu W, Meng X, Xu Q, Flower D, Li T: **Quantitative prediction of mouse class I MHC peptide binding affinity using support vector machine regression (SVR) models.** *BMC Bioinformatics* 2006, **7**:182.
13. Doytchinova I, Flower D: **Physicochemical explanation of peptide binding to HLA-A*0201 major histocompatibility complex: a three-dimensional quantitative structure-activity relationship study.** *Proteins* 2002, **48**:505–518.
14. Tian F, Yang L, Lv F, Yang Q, Zhou P: **In Silico quantitative prediction of peptides binding affinity to human MHC molecule: an intuitive quantitative structure-activity relationship approach.** *Amino Acids* 2009, **36**:535–554.
15. Cortes C, Vapnik V: **Support vector networks.** *Mach Learn* 1995, **20**:273–297.
16. Wang L: *Support Vector Machines: Theory and Applications, Volume 177 of Studies in Fuzziness and Soft Computing.* Heidelberg, Germany: Springer Berlin; 2005.
17. Nielsen M, Lund O: **NN-align. an artificial neural network-based alignment algorithm for MHC class II peptide binding prediction.** *BMC Bioinformatics* 2009, **10**:296.
18. Reche P, Glutting J, Reinherz E: **Prediction of MHC class I binding peptides using profile motifs.** *Hum Immunol* 2002, **63**:701–709.
19. Tung C, Ziehm M, Kämper A, Kohlbacher O, Ho S: **POPISK: T-Cell reactivity prediction using support vector machines and string kernels.** *BMC Bioinformatics* 2011, **12**:446.
20. Yang J, Zhang L, Zu Y, Yang JY: **Beyond sparsity: the role of l1-optimizer in pattern classification.** *Pattern Recognit* 2012, **45**:1104–1118.

doi:10.1186/1756-0381-7-23

Cite this article as: Aguilar-Bonavides *et al.*: Accurate prediction of major histocompatibility complex class II epitopes by sparse representation via ℓ_1 -minimization. *BioData Mining* 2014 **7**:23.